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DNA Disentangling by Type-2 Topoisomerases

DNA supercoil

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²Department of Molecular Virology and Microbiology Baylor College of Medicine Houston, TX 77030, USA A type-2 topoisomerase cleaves a DNA strand, passes another through the break, and then rejoins the severed ends. Because it appears that this action is as likely to increase as to decrease entanglements, the question is: how are entanglements removed? We argue that type-2 topoisomerases have evolved to act at "hooked" juxtapositions of strands (where the strands are curved toward each other). This type of juxtaposition is a natural consequence of entangled long strands. Our model accounts for the observed preference for unlinking and unknotting of short DNA plasmids by type-2 topoisomerases and well explains experimental observations.

Keywords: DNA crossovers; helix-helix juxtaposition; catenane; knot;

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Type-2 topoisomerases lower the probability of DNA linking and knotting of small plasmid DNAs by a factor of 16 to 90-fold relative to that level achieved by random DNA strand passage.¹ This finding was interpreted to mean that these enzymes prefer to disentangle, which is the required biological activity, rather than tangle DNA. Several models for how type-2 topoisomerases achieve this have been offered. Rybenkov et al. proposed that the topoisomerase tracked along two DNA helices to pinch a third.1 However, Yan *et al.* calculated that the probability that three DNA strands coincide in space was too infrequent to account for this type-2 topoisomerase activity.^{2,3} Instead, they proposed a "kinetic proofreading model" in which the enzyme, bound to one DNA helix, required two consecutive additional helix collisions to effect DNA strand passage. Vologodskii and co-workers proposed that a short region of localized high curvature, a hairpin, is created by the topoisomerase and that the DNA strand passage occurs when another DNA helix is enclosed by the hairpin.4,5 These models are locally "blind" in the sense that they do not let topoisomerase use local information already present on the substrate to determine good places for action. The hairpin, in particular, is placed by the topoisomerase at random points along the strand so that the enzyme must wait

Local structure of juxtapositions

Type-2 topoisomerases bind helix-helix juxtapositions.⁶⁻¹¹ Here, we propose that type-2 topoisomerases use the local information at the juxtaposition to distinguish an entanglement from two helices that are juxtaposed, but not entangled. To illustrate this, we depict in Figure 1 double-stranded DNA as ropes. The position of the ropes in space is determined by their center curves, K_1 and K_2 . Let P_1 , P_2 denote the points on the curves K_1 , K_2 , respectively, such that $|P_1 - P_2|$ is mini-mized. Let p_{12} denote $P_1 - P_2$, and $p_{21} = P_2 - P_1$ denote the separation vectors for P_1 and P_2 . The juxtaposition exists when $|p_{12}| < c$, for some pre-determined constant c, let us say roughly equal the diameter of a type-2 topoisomerase, ~ 100 Å. Let T_1 , T_2 denote unit tangent vectors to K_1 , K_2 at P_1 , P_2 , respectively. Let r_1 , r_2 denote the curvature vectors at P_1 and P_2 .

We could classify the juxtapositions by the triple of vectors T_1 T_2 and the separation vector, which would allow us to distinguish between right and left hand. The categories could be further subdi-vided by considering the magnitudes of the curva-tures and subdivisions of the angles between the tangents. Because DNA contains directional read-out, that information is also present at a juxtaposi-tion. We can also consider the time derivatives of the vector sets shown in Figure 1E and denote

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until strands are brought into juxtaposition to then act.



Figure 1. How strands juxtapose. We show four cases. Hooked juxtapositions are cases A and E, where the curves are curved toward each other $(r_1 \cdot p_{21} > 0 \text{ and } r_2 \cdot p_{12} > 0)$ and B, where one curve encloses the other $(r_1 \cdot p_{21} < 0 \text{ and } r_2 \cdot p_{12} > 0 \text{ and } r_1 > r_2)$. Free juxtapositions are case C, where the curves curve away $(r_1 \cdot p_{21} < 0 \text{ and } r_2 \cdot p_{12} < 0)$, and case D, where the inner curve has the greater curvature $(r_1 \cdot p_{21} > 0 \text{ and } r_2 \cdot p_{12} < 0 \text{ and } r_1 > r_2)$. Through most of this work we are concerned with cases A and C. F, Supercoiled links tend to have hooked juxtapositions. G, The most exposed sections of loops tend to be those that curve away from an arbitrary path of approach, so unlinked loops tend to have free juxtapositions. H, Two tangled loops being pulled apart, creating hooked juxtapositions.

them P_1t , P_2t , $p_{12}t$, etc. Each of these has geometrical interpretation. $p_{12}t$, for example, can measure the rate at which the closest points tend to come together or separate. If $p_{12}t$ is small, the juxtaposition is persistent. If P_1t is large, but P_2t and

 $p_{12}t$ are small, we could imagine that K_1 is locally sliding past K_2 , an axial motion. Some sort of pulling force or random motion could cause $r_1 t > 0$ and $r_2 t > 0$.

Methods for recognizing local structure

One way that type-2 topoisomerases might distinguish juxtapositions would be if, built in to the enzyme, was a template into which the juxtaposition fit or not. Alternatively, curvature could be read from the bending strain on the helical structure. Right and left-handed juxtapositions lie upon each other differently,10 which provides a way for topoisomerases to distinguish handedness.^{12,13} Because type-2 topoisomerases have preferred DNA cleavage sequences, it is likely that DNA read-out influences how these enzymes distinguish juxtapositions. Time may come into play. If a type-2 topoisomerase acts with some likelihood on any juxtaposition, then those juxtapositions with greater persistence may be preferentially recognized. With time, fluctuations could place the DNA strands in correct position for the topoisomerase to fit a template. This would happen quickly for juxtapositions that are already near the template shape, slowly for ones that are not. Curvature and persistence are connected through two routes. First, hooked juxtapositions have greater local constraints than free juxtapositions. A simplistic analysis: assume that the two strands are relatively stiff and consider the

local rigid motions of K_1 , holding K_2 fixed. A basis for these motions is the positive and negative translations in the p_{12} , T_1 and B_1 directions (where B_1 is the binormal at P_1), and the three rotations about these vectors (assume the origin is at P_1). For the hooked juxtaposition in Figure 1A, the motion is constrained in each translational direc-tion except the negative p_{12} , and in two of the three rotations. In the free juxtaposition case in Figure 1C, the motion is constrained only in the negative p_{12} direction of the six translations and in only one of the rotations, and even that is a large angle constraint. Similar considerations show that the hooked in Figure 1B is more constrained than the free in Figure 1D, but the difference is less than for A and C (Figure 1).

Second, it is generally held that two DNA helices must overcome electrostatic repulsion to create a juxtaposition. Consider the potential energy of two like charged strands, spreading the charge out uniformly along the strands (assume unit charge density). We calculate only the effect of each strand upon the other, and not the effect of the strand upon itself (the self-potential of a charged string is well known to have a logarithmic singularity.14 The assumption is that the bonds along the strand locally counteract the self-potential. Here, strands are bent at a $\pi/2$ angle, and moved toward one another by rigid translation (Figure 2A). The maximum energy takes place when p > 0, and decreases through p = 0, so that once the strands are close enough, bringing them even closer is energetically favored (Figure 2B). Therefore, hooked juxtapositions are locally attracting (or at least not repelling), and free juxtapositions are repelling.

Similar considerations hold for the angle, α ,



Figure 2. Effect of charge on strand juxtapositions. In B, the x-axis is the separation, p, depicted in A, the y-axis is the electrostatic potential *E* of the strands upon each other. For this computation, the edges are of length square-root 2, the charge density along the edges is constant 1, and the graph is of the integral of the inverse distance between pairs of points on the edges, where the distance p is the distance between the central vertices of the two angled filaments. The graph was computed by the software Mathematica, the complete code is: $x1[t_{-}] = t$; $y1[t_{-}] = t$; $z1[t_{-}] = 0.0$; $K1[t_{-}]:=\{x1[t_{-}],y1[t_{-}],z1[t_{-}]\}; \quad x2[s_{-}]:=0.0; \quad y2[s_{-}]:=-s+m; \quad z2[s_{-}]:=s; \quad K2[s_{-}]:=\{x2[s_{-}],y2[s_{-}],z2[s_{-}]\}; \quad CHPL[v_{-}]:=0.0; \quad y2[s_{-}]:=0.0; \quad y2[s_{-$ NIntegrate[2/(((K1[t] - K2[s]))(K1[t] - K2[s]))^5, [t, 0, 1], [s, 0, 1]]; Plot[CHPL[v], [m, -4, 4]]. Note that this is the contribution from the interaction of one edge with both edges of the other filament. For the full cross-potential the numerator of the integrand should be 4, which of course does not change the shape of the curve. A, was rendered with Pov-Ray. For C, discrete charges were uniformly spread along the filaments and all interactions between charges were calculated. The endpoints of the strands form a regular tetrahedron and $\alpha = \pi/2$, where α is the angle between the tangents at the juxtaposition. The software Knotplot was employed for both the calculation and the rendering.

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between the tangents. In Figure 2C is depicted the result of a numerical computation of the minimum electrostatic energy conformation of two flexible strands that are joined at their center points. If p = 0, hooked and free juxtapositions are indistinguishable. The conformation is non-planar and so has non-zero curvature at the midpoint of the strands, an additional connection between curvature and juxtapositions.

How global topology is expressed in local structure

392 A thought experiment illustrates how type-2 393 topoisomerases may act. Imagine N (large number) 394 exact (perfectly round) circles moving by random 395 continuous motion within a finite volume. Employ 396 a rule, rule H: if the juxtaposition is hooked, then 397 the strands are passed through one another; other-398 wise they are not. Because we have exact circles, 399 rule H will not link any pair and, as long as the 400 circles are free to move, it will eventually unlink 401 all linked pairs. So the long-run behavior is N402 unlinked circles: trivial topology. In contrast, rule 403 F passes DNA strands only in free juxtapositions. 404 There is no unlinking, so in the long run every 405 circle is linked. Rules H and F use local juxta-406 position information to arrive at opposite topologi-407 cal outcomes. This result is dependent on the 408 length and rigidity of the circles. That is, as long 409 as the loops are perfect geometric circles, the local 410 geometry completely determines the topology. 411 However, DNA is an elastic filament with bending 412 energy. So increasing length adds geometric flexi-413 bility, the loops move further and further away 414 from perfect circle conformations. We can imagine 415 the opposite end of the spectrum from the perfect 416 circle case: very long loops packed at a high den-417 sity and mixed about one another, think of a bowl of spaghetti. Because the loops are very long, and 418 419 intertwine with one another, some rule H crossings 420 will now create links. Either rules H or F, applied 421 to a system of arbitrarily long chains in constant 422 density, would tend to the same order of topologi-423 cal complexity proportional to $L^{4/3}$, where \hat{L} is the 424 total length of the system.15 Consider an inter-425 mediate case: random closed loops of a fixed 426 length, sparsely distributed. Isolated closed loops 427 have fewer rule H than rule F accessibility, mean-428 ing that when two isolated loops come into contact, 429 the juxtapositions are more likely to be free than 430 hooked. Rule H will, in this case, decrease linking, 431 though not to the extent it does for perfect circles. 432 In general, the effectiveness of a rule that depends 433 upon local information will depend upon both the 434 length and the density of the DNA strands.

⁴³⁵ Let S1 be the smallest sphere containing loop K_1 , ⁴³⁶ S2 the smallest sphere containing K_2 . Generically, ⁴³⁷ S1(S2) is determined by four points along K_1 . At ⁴³⁸ these points the curvature vector points inside ⁴³⁹ S1(S2). Assume K_1 and K_2 are not linked, and ⁴⁴⁰ brought near one another. The points most likely ⁴⁴¹ to be brought into juxtaposition are near these DNA Disentangling by Type-2 Topoisomerases

442 external points (see Figure 1F). However, such jux-443 tapositions cannot be hooked. More generally, 444 place two unit spheres randomly in a large box 445 and compute the conditional probability that, if 446 they intersect, how far apart are their centers. 447 Shallow intersections (distance between centers between 1/2 and 1) are roughly sevenfold more 448 449 likely than deep (distance between 0 and 1/2) 450 ones. However, shallow intersections tend to give free juxtapositions of unlinked loops. In general, if 451 452 we approach the loop from an arbitrary direction, 453 then at the points most likely to hit first, the strand 454 is curved away from the direction of approach.

455 Therefore, we define rule H (in)accessibility. 456 A tightly wound ball of string is perfectly rule H 457 inaccessible: from all directions of approach the 458 string curves away at the point of first incidence. 459 Two compact globules are very unlikely to have 460 hooked juxtapositions when they first meet. On 461 the other hand, the external loops of any conformation provide opportunities for rule F crossing 462 463 change. In contrast, let K_1 and K_2 be linked. Under 464 random motion, their centers drift apart. This eventually provides a kind of "pulling apart" force, which results in hooked juxtapositions. 465 466 A rule H move, or a series of rule H moves, will 467 468 unlink them.

469 The analysis for untying knots is similar to that 470 for unlinking catenanes. The principal difference 471 is that knots are built out of several partial loops. 472 A typical example is pictured in Figure 3. If the 473 DNA were a phantom chain (allowing every strand 474 to pass through), then the labeled crossing change 475 would create a trefoil, the DNA strand behind 476 must pass through the loop. The loop has a de 477 facto inside and outside. Getting from the outside 478 to the inside cannot be a rule H move. Thus, by rule H, knots are not likely to be tied, especially 479 for the short lengths of a plasmid. On the other 480 hand, if we start with a trefoil knot, then a bend in the strand passing through the loop, even a slight one, creates a hooked juxtaposition, so rule H unties the knot. This effect is more pronounced in



Figure 3. Random strand passage would generate a knot. A strand crossing change at the indicated position would create a trefoil, but this is unlikely to happen by rule H.

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knots than in catenanes of the same length, because the subarcs of the knot are the loops and these loops are even shorter in length than the catenane loops. As with links, the proportion of hooked to free juxtapositions in a knot depends upon strand length and density.

In numerical work, it has been observed that long, random chains tend to localize a good portion of their knotting.^{16,17} By inspection, we observe that these conformations usually have outer arcs bent around inner lengths of lesser curvature. These are hooked juxtapositions by our definition.¹⁵ These conformations are not generic in our sense because they have continuous families of closest pairs of points, but they help illustrate the role of the length constraints.

Additional experimental results and our interpretations

525 Type-2 topoisomerases preferentially recognize 526 and remove the biologically problematic DNA 527 topological obstructions, knots and catenanes, 528 over biologically required DNA supercoils.18-21 529 DNA supercoiling increases the effectiveness of 530 type-2 topoisomerases in unlinking catenanes, and has a lesser or no effect on knots.^{20–23} These results 531 532 are not reconciled by any existing model. Our pos-533 tulate that type-2 topoisomerase follow rule H 534 explains these data. For a supercoiled closed loop, 535 an approach from any direction first hits a subarc curving away from the direction of approach, so 536 537 that the probability of a hooked juxtaposition 538 occurring between two loops is nearly 0. If the 539 supercoiled loops are linked already, a juxta-540 position is likely to be hooked (see Figure 1F). If a 541 minimum curvature threshold exists for rule H 542 action, then supercoiling makes this more likely to 543 be reached. The case of a supercoiled knot is 544 different. Because the strands in a knot node are 545 already interacting with each other, supercoiling 546 would have a lesser effect because these juxta-547 positions are already rule H friendly. Thus, rule H 548 will unlink supercoiled catenanes more efficiently 549 than it will supercoiled knots. Because the curva-550 ture in the juxtaposition between the loops shown 551 in Figure 1F is greater than that in the juxta-552 positions created by supercoiling, a type-2 topo-553 isomerase would unlink or unknot before it would 554 relax supercoils.

555 To model a system such as that in the Rybenkov 556 *et al.* experiment,¹ we reason dynamically, and 557 assume that the system will equilibrate at 558 M = L/U, where *L* is the rate of linking, and *U* is 559 the rate of unlinking. For example, for rule H on 560 perfect circles, L = 0 and U > 0, so the limit is 0. 561 For rule F on perfect circles, L > 0, U = 0, so the 562 limit is M = infinity (all loops are linked). Above 563 we observed that in the catenane case supercoiling 564 decreases L and increases U. The experimental 565 system used in which DNA loops were opened 566 and closed with some frequency¹ should be 567 roughly equivalent to a rule where some fixed per-

568 centage of all juxtapositions allows DNA strand 569 passage (so rule F pass throughs are allowed). 570 When a type-2 topoisomerase was introduced, the 571 equilibrium was lowered. A type-2 topoisomerase 572 has a lower ratio of L to U than a "blind" system 573 that cannot distinguish between types of juxta-574 positions. Let L_b and U_b be the constants for this 575 blind system, L_{to} and U_{to} be the constants for a sys-576 tem governed by rule H. Then the equilibrium is 577 $M = (L_{\rm b} + L_{\rm to})/(U_{\rm b} + U_{\rm to})$. Many factors determine 578 L and U. At least two can create a difference in the 579 speeds of the reactions. The loops may open and 580 close at speeds differing from type-2 topo-581 isomerase reaction speed. Also, the opening and 582 closing of one of the loops in this system occurs at 583 only one locus along the DNA strand, but topoi-584 somerase should be free to act anywhere along the 585 loop. Therefore, even if the gate opened and closed 586 with speed comparable to enzyme action, the effec-587 tive speed would be reduced by the proportion of 588 the size of the gate to the arclength of the loop. 589 Hence, the natural assumption that the equilibrium 590 of the combined system is close to $L_{\rm to}/U_{\rm to}$. For 591 knots, we let K denote the knotting constant and 592 *U* the unknotting constant. Our analysis gives a 593 prediction for the dependencies of 594 $(L_b/U_b)/(L_{to}/U_{to})$ and $(K_b/U_b)/(K_{to}/U_{to})$ on loop 595 length, which is depicted schematically in Figure 4A. 596

597 We can relate Figure 4A to the experimental 598 evidence:¹ a 16-fold reduction (relative to what 599 was termed "topological equilibrium") in linking 600 in loops of length 10 kb (thus 20 kb total length), a 601 40-fold reduction in knotting in loops of length 602 10 kb, and an 800-fold reduction in knotting in 603 loops of length 7 kb (Figure 4B). As illustrated in 604 Figure 4A, there is a vertical asymptote at length = 605 C. Our discussion of rule H inaccessibility gave a rough estimate of D = 8 for longer loops in low-606 density situations. The limiting value D depends 607 on density, but we can estimate that under similar 608 609 experimental conditions, at 50 kb or 100 kb, the 610 ratio would be nearly *D*.

611 Rybenkov et al. reported that type-2 topoisome-612 rase reduces the variance of the linking number 613 distribution for a plasmid DNA, meaning that the enzyme creates a distribution of supercoiling more 614 615 tightly clustered about 0 than a random generation method.¹ We interpret these findings to mean that 616 617 greater supercoiling, on average, creates more 618 hooked juxtapositions with higher curvature (because length is fixed), so the loops with greater 619 620 supercoiling are more likely to be acted upon by 621 rule H.

622 In the Rybenkov experiments,¹ in the measure-623 ment of entanglement as a function of enzyme con-624 centration, it was seen that entanglement declines 625 with increasing concentration, reaches a minimum, 626 then increases with increasing concentration. We 627 can explain this as follows. In the experiments, the loops are opening and closing at some rate. When 628 629 the enzyme concentration is very low, the gates, 630 on average, create entanglements faster than the



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694 Figure 4. Length dependence of 695 knotting and linking. A, The length is the length of an individual loop. 696 We assume that the strands have 697 some elastic resistance to bending, 698 some thickness, and that the overall 699 density is low. For linking, the 700 asymptote at length = C: as the cir-701 cles get shorter, they must become 702 more like perfect circles because of 703 the bending constraints. However, 704 rule H is perfect on perfect circles, 705 so $L_{to}/U_{to} = 0$, but $L_b/U_b > 0$, hence the vertical asymptote. The 706 horizontal asymptote at 707 $(L_{\rm b}/U_{\rm b})/(L_{\rm to}/U_{\rm to}) = D^{-}$ as length 708 tends to infinity. Given rule H inac-709 cessibility, we have that even very 710 long loops, if density is sufficiently 711 low, are less likely to be linked 712 under rule H than by arbitrary pas-713 sage, hence the limit at D > 1. If 714 instead we allow the density to 715 increase as we increase length, so the loops become both very long 716 and completely intermingled, then 717 we conjecture that D = 1 and topoi-718 somerase cannot lower the equili-719 brium. We expect different values 720 for *C* and *D* for knots. The asymp-721 tote at length = C: as the length 722 gets small, the ratio of free to 723 hooked juxtapositions in knots 724 goes to zero, knots are formed by 725 subarcs of the circle that themselves, because of the constraints, 726

behave like perfect circles in that they are rule H inaccessible. (In tight conformations the curvature of one strand is 664 727 greater than that of the strand passing through it). The horizontal asymptote at $(K_b/U_b)/(K_{to}/U_{to}) = D$ as length 665 728 tends to infinity: this case is similar to the linking analysis, in that the density is an important factor. The mathematical 666 729 theory here is incomplete, but models based on random walks in unconstrained spaces show that chains tend to localize at least some of their knotting and tangling.^{16,17} The curvature is likely to be high at the localization, ripe for 667 730 668 731 rule H action. In a localized knot component, the effective length of that part of the strand is small, so we effectively 669 732 are close to the value length = C, so we expect that type-2 topoisomerase would be much more likely to remove a 670 733 localized knot that to create one. On the other hand, it has been shown that in unconstrained random walks there is 671 at least some global knotting, so the determination of D in this model is an open question. An unconstrained random 734 672 walk is only a model for DNA, and clearly has some deficiencies as a model for DNA in the cell, where the packing 735 is not random and there are severe volumetric constraints. Again, the mathematical theory is lacking, but it is plausible 673 736 that a volume constraint would give a greater percentage of global knotting, thus reducing D for random chains. 674 737 B, Data are from Rybenkov et al^{1} The fold reduction is relative to the amount of linking (or knotting) seen in 675 738 equilibrium without type-2 topoisomerase. 676 739

678 enzyme can act, thus near the equilibrium value of 679 entanglement. As the concentration increases, the 680 enzyme can, on average, act faster, and so detangle 681 the system. To see what happens when concen-682 tration increases beyond biological levels, we con-683 sider further the connection between hooked 684 juxtapositions and persistence. If the loops are 685 long enough, then two unlinked loops may meet 686 in a hooked juxtaposition. Our argument is not 687 that hooked juxtapositions are impossible for 688 unlinked loops, just that they are less likely than 689 free ones. Because they are unlinked, this juxta-690 position is not likely to be as persistent as a hooked 691 juxtaposition of linked loops. The salient point is 692 that there is information in the length of time juxta-693 position persists. If the concentration of the

741 enzyme is very high, then all hooked juxtapositions, both those that would have been tran-742 743 sient and those that would have been persistent, are acted on immediately. So this persistence infor-744 745 mation is lost, as a result of which the ratio of per-746 sistent to transient hooked juxtapositions acted 747 upon is given by the ratio of the rates at which the 748 respective juxtapositions are formed. If, on the 749 other hand, the average time it takes the enzyme 750 to act is long (lower concentration), then the proportion of transient to persistent acted upon now 751 752 depends on the length of time the juxtapositions 753 persist. We imagine that this phenomenon also happens in the cell, that for this reason too much 754 755 enzyme could actually slow disentanglement and 756 therefore cell division.

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